

Diet and female fertility: doctor, what should I eat?

Yu-Han Chiu, M.D., Sc.D.,^a Jorge E. Chavarro, M.D., ScD,^{a,b,c} and Irene Souter, M.D.^d

^a Department of Nutrition and ^b Department of Epidemiology, Harvard T.H. Chan School of Public Health; ^c Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School; and ^d Division of Reproductive Endocrinology and Infertility, Department of Obstetrics and Gynecology, Harvard Medical School, Massachusetts General Hospital Fertility Center, Boston, Massachusetts

Fecundity is the capacity to produce offspring. Identifying dietary factors that influence human fecundity is of major clinical and public health significance. This review focuses on the evidence from epidemiologic literature for the relationships between key nutritional factors and female reproductive potential. According to existing data, women trying to achieve pregnancy are encouraged to increase consumption of whole grains, omega-3 fatty acids, fish, and soy and to reduce consumption of *trans* fats and red meat. In addition, a daily multivitamin that contains folic acid before and during pregnancy may not only prevent birth defects, but also improve the chance of achieving and maintaining a pregnancy. In contrast, there is limited evidence supporting an association between vitamin D and human fecundity outcomes despite promising evidence from nonhuman studies. Questions for future research included the roles of other types of fat (especially omega-6 and monounsaturated fats) and protein (especially white meat and seafood) on female fertility; particular attention should also be paid to exposure to environmental contaminants in foods. Although much work remains, this review accrued best available evidence to provide practical dietary recommendations for women trying to conceive. (Fertil Steril® 2018;110:560–9. ©2018 by American Society for Reproductive Medicine.)

Key Words: Diet, dietary patterns, micronutrients, macronutrients, female fertility

Discuss: You can discuss this article with its authors and other readers at <https://www.fertstertdialog.com/users/16110-fertility-and-sterility/posts/33449-26262>

It is estimated that infertility affects 15.5% of reproductive-age women in the United States (1), and 30% of pregnancies are lost after implantation (2). Although assisted reproductive technologies (ART) become a common treatment choice, because of the financial and emotional challenges associated with ART, emerging scientific efforts focus on the identification of modifiable factors, such as diet and lifestyles, that may affect fertility. Dietary factors have been implicated in the pathology of multiple disorders (3–6), and the idea that dietary changes may boost fertility appears to be promising.

Human fecundity is difficult to assess directly. Therefore, most researchers rely on proxy measures,

such as assessment of time to pregnancy (a shorter time indicating a higher fecundity) and whether a pregnancy, pregnancy loss, or live birth occurs among pregnancy planners or women undergoing ART. Other commonly used fecundity proxies include medically determined causes of infertility, reproductive hormonal profiles and menstrual irregularities (to assess ovulatory function), and ovarian antral follicle counts as well as serum levels of antimüllerian hormone (to assess ovarian reserve). In this review, we summarize the evidence from human studies relating diet to these markers of fecundity for the purpose of providing a tool to counsel patients trying to achieve pregnancy.

MICRONUTRIENTS

Folic Acid

Folate, involved in the synthesis of DNA (7), is crucial in gametogenesis, fertilization, and pregnancy (8, 9). Therefore, folate (natural form of vitamin B9) or folic acid (synthetic form of vitamin B9) may play an important role in human reproduction.

Folic acid and the risk for spontaneous abortion. Since the early 1990, the U.S. Public Health Service and Centers for Disease Control and Prevention have recommended that all women of childbearing age take a daily supplement containing 0.4–0.8 mg folic acid to prevent neural tube defects (10). In the mid-1990s, controversy over the safety of folic acid supplementation arose when three studies reported increased spontaneous abortion (SAB) rates among folic acid users (11–13). The validity of these findings was later challenged on methodologic grounds (14, 15), and in the most recent Cochrane review (16), on the basis of three randomized trials

Received May 8, 2018; accepted May 23, 2018.

J.E.C. and I.S. should be considered similar in author order.

Y.-H.C. has nothing to disclose. J.E.C. has nothing to disclose. I.S. has nothing to disclose.

Supported by grants P30ES000002, R01ES009718, R01ES022955, and P30DK046200 from the National Institutes of Health.

Reprint requests: Irene Souter, M.D., Massachusetts General Hospital Fertility Center, Yawkey 10-A, 55 Fruit Street, Boston, MA 02114 (E-mail: isouter@partners.org).

Fertility and Sterility® Vol. 110, No. 4, September 2018 0015-0282/\$36.00

Copyright ©2018 Published by Elsevier Inc. on behalf of the American Society for Reproductive Medicine

<https://doi.org/10.1016/j.fertnstert.2018.05.027>

(11, 17, 18), daily folic acid (0.8 mg in one study and 4 mg in two studies) plus multivitamin supplementation before and during pregnancy did not increase SAB rates among users versus nonusers. Similarly, a large population-based study in China found no increased risk for SAB among daily consumers of folic acid (19), and a Brazilian multicenter trial reported no difference in SAB rates between high and low folic acid supplementation groups (0.4 vs. 4 mg/d) (20). Interestingly, recent data from observational studies, including a large prospective cohort of healthy women in the Nurse's Healthy Study II (NHS-II), suggested a reduced SAB risk among women using folic acid before or during early pregnancy, particularly at intake levels well above those recommended for the prevention of neural tube defects (21–23).

Folic acid, fecundity, and ovulatory infertility. The associations between folic supplementation and infertility have also been examined in three prospective cohort studies, which in general suggested a protective effect. Specifically, among women from the NHS-II study, multivitamin users had approximately one-third lower risk of developing ovulatory infertility compared to nonusers, and folic acids appeared to explain most of this association (24). Similarly, folate intake was related to a lower frequency of sporadic anovulation in a prospective cohort of young healthy women (the Biocycle study) (25) and to a shorter time to pregnancy among pregnancy planners in a large Danish cohort (26).

Folic acid and ART outcomes. Studies among subfertile women generally suggest a favorable effect of folate supplementation on ART outcomes. In a small randomized controlled trial (RCT) of subfertile women, women who took a daily multivitamin (containing 0.4 mg folic acid) had 16% higher probability of pregnancy than women in the placebo group (27). In addition, in two studies, the *MTHFR* 677T allele mutation (leading to lower *MTHFR* enzyme activity and lower serum folate levels) was associated with poor ovarian response, fewer retrieved oocytes (28), and lower granulosa-cell E_2 production than the wild-type allele (29). Furthermore, in a prospective ART cohort in Boston (EARTH study), women consuming >0.8 mg/d folate compared with those consuming <0.4 mg/d, before conception, had a higher probability of live birth (30). In this same study, higher serum levels of folate and vitamin B₁₂ measured during the stimulation phase of the cycle were associated with a higher probability of live birth (31). Nonetheless, the results from three European cohort studies of folate and ART outcomes did not show similar benefits (32–34). The latter results, however, should be interpreted with caution considering they excluded women who failed before embryo transfer. If folates affect early ART outcomes, as suggested by some studies (30, 35–37), then excluding these women could bias the results toward the null.

Summary. Overall, current evidence generally supports folic acid supplementation before and during pregnancy, because it appears that folate is not associated with higher risks of SAB but may instead improve a woman's chance of achieving and maintaining a pregnancy. Benefits seem to appear at intakes above those recommended for the prevention of neural tube defects, but trials testing these doses in relation to fertility are lacking.

Vitamin D

Accruing literature suggests that vitamin D may modulate reproductive processes. Vitamin D receptors are widely distributed across the reproductive system, including ovaries, uterus, and endometrium (38). Animal studies have shown that female rodents fed a vitamin D deficient diets, as well as knockouts for vitamin D receptors and 1α -hydroxylase (enzyme responsible for converting circulating 25-hydroxy vitamin D₃ [25(OH)D] to its biologically active form) have reduced fertility (39–43). Furthermore, vitamin D stimulates ovarian steroidogenesis, promotes follicular maturation, and regulates *HOXA10* expression (involved in successful implantation) (44, 45), and its deficiency may be involved in the pathogenesis of polycystic ovary syndrome (PCOS).

Vitamin D and reproductive outcomes. Despite a promising role of vitamin D in reproduction in nonhuman animal studies, studies evaluating the relation between vitamin D and fecundity in healthy human populations generally show no strong associations. Among women participating in the NHS-II study, vitamin D intake was unrelated to anovulatory infertility (46). Similarly, vitamin D concentrations were not associated with either the overall probability of conception (among healthy Danish women) (47) or conception in less than 1 year (among Italian women undergoing routine aneuploidy screening) (48). Furthermore, a meta-analysis (49) of 10,630 pregnant women from five cohort studies (50–54) revealed no association between low 25(OH)D levels and SAB risks, although extremely low levels (<20 ng/mL) were associated with increased early SAB risk in a subgroup analysis of two studies (50, 53).

Vitamin D and ART outcomes. Results concerning a possible role of vitamin D on ART outcomes are inconsistent. In a recent meta-analysis of 11 cohort studies (five prospective [55–59] and six retrospective [60–65]) of women undergoing ART (66), Chu et al. found that women replete in vitamin D, compared with women with either deficient or insufficient vitamin D levels, had higher probability of clinical pregnancy and live birth. No association of vitamin D with probability of miscarriage was noted (66). Similarly, a post hoc analysis of an RCT in PCOS patients found that serum 25(OH)D <30 versus >30 ng/mL was associated with lower live birth rates (67). In contrast, three small observational studies, not included in this meta-analysis, found no association between serum or follicular fluid vitamin D concentrations and ART outcomes (68–70). Furthermore, findings from two small RCTs did not support the administration of vitamin D to improve pregnancy outcomes (71, 72). Neither weekly supplementation of 50,000 IU vitamin D for 6–8 weeks to deficient women (71) nor administration of megadose vitamin D (300,000 IU) to women with PCOS improved reproductive outcomes (72). In the latter, a significant increase in endometrial thickness was noted but did not translate to a significantly higher probability of pregnancy (72).

Summary. Despite there being promising mechanisms through which vitamin D can affect reproduction, evidence from epidemiologic studies remains inconclusive, though

suggestive that serum levels in the deficiency range are related to worse outcomes in ART. Larger RCTs are required to examine the effect of vitamin D supplementation on fertility, as well as to determine who will benefit from the supplementation and at what doses.

MACRONUTRIENTS

Carbohydrates

Both quality and quantity of dietary carbohydrates influence glucose homeostasis and insulin sensitivity (73), which may in turn influence ovarian androgen production and ovarian function. The common indicators of carbohydrate quality include glycemic index (an index to indicate the effect of carbohydrate on blood glucose), glycemic load (a product of glycemic index and amount of carbohydrates), the extent to which carbohydrate have been refined (whole grains vs. refined grains), and amount of dietary fiber.

Glycemic load and reproductive outcomes. In NHS-II, both total carbohydrate consumption and glycemic load were associated with higher risks of ovulatory infertility (74). Consistently with this finding, several studies showed that women with PCOS more often exhibit a dietary pattern marked by a greater consumption of high-glycemic-index foods compared with normoandrogenic women (75–77). Reduction in dietary carbohydrates among PCOS women improved insulin sensitivity (78–80) and decreased circulating testosterone levels (79), potentially enhancing ovulatory function. Nonetheless, among healthy regularly menstruating women, dietary carbohydrate intake was unrelated to androgens and relevant hormones (testosterone, antimüllerian hormone, insulin) (81), possibly owing to a relatively narrow range of hormone concentrations in the healthy population.

Whole grains and ART outcomes. Whole grains and constituents of whole grains (including phytic acid, vitamins, and selenium), which have antioxidant antiinflammatory properties and beneficial effects on glucose metabolism, may potentially boost fertility because insulin resistance and oxidative damage have been implicated in the pathogenesis of subfertility (82). Furthermore, lignan (83), the hormonally active compound in whole grains, through its proestrogenic and antiestrogenic effects, may exert reproductive benefits. A prospective study of women attending a fertility clinic (EARTH study) showed that higher preconception intake of whole grain was associated with higher probability of live birth (84).

Dietary fiber and reproductive and ART outcomes. In a couple of clinical trials of premenopausal women, high-fiber/low-fat diet was associated with reduced estrogen levels (85–89), presumably because high-fiber diet decreases fecal β -glucuronidase activity, thus decreasing reabsorption of estrogen (90). Similarly, in a study of regularly menstruating women (the Biocycle study), a diet high in fiber was associated with decreased E_2 levels (91). Regarding its association with ovulation, a diet rich in fiber was associated with an increased risk of anovulation in one study (91) but was unrelated to ovulatory infertility in the long run in another (NHS-II) (74). Furthermore, total fiber intake was unrelated to ART success among women undergoing fertility treatments (EARTH

Study), but intake of bran was responsible for the positive association between whole grains and live birth rates described above (84).

Summary. Current evidence, though limited, suggests that a diet low in glycemic load and containing greater amounts of whole grains may benefit fecundity, and that a diet rich in fiber may reduce estrogen levels but may not affect either the risk of infertility or ART outcomes.

Fatty Acids

Fatty acids, commonly classified as saturated (SFA), monounsaturated (MUFA), and polyunsaturated (PUFA; including ω -3 PUFA and ω -6 PUFA), may play important roles on reproductive function through myriad pathways. For one, fatty acids are used as energy substrates during oocyte maturation and early embryo development (92), and they serve as critical precursors for a variety of substrates (e.g., prostaglandins and steroid hormones) playing a vital role in implantation and pregnancy maintenance (93). On the other hand, *trans* fatty acids increase insulin resistance (94), thus adversely affecting the ovulation process (95).

***Trans* fat in ovulatory infertility, endometriosis, and fecundity.** In a cohort of regularly menstruating healthy women (the Biocycle study), *trans* fat was unrelated to either hormone concentrations or risks of anovulation (96). However, in the NHS-II study, consuming *trans* fat instead of carbohydrate or other unsaturated fat was associated with higher risks of ovulatory infertility (97) and laparoscopically confirmed endometriosis (98). Furthermore, *trans* fat intake was associated with reduced fecundability in a large North American preconception cohort (PRESTO study) (99). However, no such association was observed in a similar Danish preconception cohort (Snart Foraeldre study), where *trans* fat intake was low (99).

ω -3 PUFA and fecundity, ovulatory infertility, and ovarian aging. In the Biocycle study, increased long-chain ω -3 PUFA intake was associated with increases in luteal progesterone concentration; furthermore, docosapentaenoic acid (a type of long-chain ω -3 PUFA) was associated with increased total E_2 and lower risk of anovulation (96). Although no association was identified between ω -3 PUFA and ovulatory infertility in the NHS-II study (97), an association between greater consumption of ω -3 PUFA and a lower risk of endometriosis was noted (98). Furthermore, ω -3 PUFA was associated with higher fecundability among women who did not use fish oil supplements in the PRESTO study (U.S. and Canada), although no association was found in the Snart Foraeldre cohort (Denmark), among whom baseline intake was significantly higher (99).

Regarding potential effects on ovarian reserve, dietary administration of ω -3 PUFA decreased serum FSH levels in normal-weight but not in obese women with normal ovarian reserve (100), which is consistent with murine data whereby higher dietary ω -3 PUFA delayed ovarian aging (101).

ω -3 PUFA and ART outcomes. The favorable associations between ω -3 PUFA and reproductive end points were identified in ART settings as well. The EARTH study reported that

for every 1% increase in serum long-chain ω -3 PUFA levels, the probability of clinical pregnancy and live birth increased by 8% (102). In another study, serum levels of eicosapentaenoic acid (a long-chain ω -3 PUFA) were significantly higher in pregnant than in nonpregnant patients (103). In addition, higher preconception ω -3 PUFA intake was associated with better embryo morphology, despite being inversely associated with response to ovarian stimulation (104). Furthermore, although Jungheim et al. found, in a small ART cohort of 91 women, that serum α -linolenic acid (ALA; a type of ω -3 PUFA) concentrations were associated with decreased chances of pregnancy (105), in a subsequent analysis of a larger number of women ($n = 200$), the authors reported no association between any individual PUFA and ART outcomes (106). Finally, despite Jungheim et al. finding a positive association between the ratio of linoleic acid (LA) to ALA with higher implantation and pregnancy rates (106), that finding was not replicated in the EARTH cohort (102).

ω -6 PUFA and ART outcomes. Results concerning ω -6 PUFA were less consistent across studies. In a small study of overweight and obese women undergoing ART, preconception intake of ω -6 PUFA (especially LA) was associated with improved pregnancy rates (107). In another, though, follicular fluid levels of LA negatively correlated with metaphase II oocytes. Similarly, in the EARTH study, serum LA levels were inversely associated with total oocyte and metaphase II oocyte yield, but this association did not translate into differences in implantation, clinical pregnancy, or live birth (102).

Summary. Taken together, higher ω -3 PUFA and lower *trans* fatty acid intake may enhance female fertility. The effect of other fatty acids, including ω -6 PUFA, SFA, and MUFA, on fertility is less clear.

Protein

The daily recommended dietary allowance of protein for a healthy adult with minimal physical activity is 0.8 g per kg of body weight. Recommendations on protein intake (amount and type) for women attempting conception or undergoing fertility treatments do not exist, and the controversy surrounding the evidence on animal and dairy protein as well as soy and phytoestrogens remains. Concerns have arisen because of the potential for contamination of dietary protein sources by pesticides and endocrine-disrupting chemicals (108–110) and the presence in them of measurable amounts of steroid hormones and growth factors (111–114) that when absorbed may alter reproductive outcomes. The mechanisms involved are unknown, but absorbed contaminants may alter serum levels of growth factors or adversely affect either the hypothalamic-pituitary-gonadal axis or the oocyte and its local supporting environment through modifications of gene expression and neuroendocrine signaling (115–120).

Dairy and ovarian reserve. Earlier studies in rodents showed that in galactose-exposed mice puberty was delayed, response to gonadotropins was blunted, and follicular reserve was decreased, suggesting that dairy's high galactose content might have a negative impact on ovarian physiology (121).

In human populations with higher per-capita consumption of milk, the decline in fertility with aging was steeper and fertility at older ages was lower (122). Similarly, higher dairy protein intake, and not total protein intake, was associated with lower ovarian antral follicle counts among women presenting for infertility treatment (123). Although these findings suggest a possible negative impact of dairy on ovarian reserve, studies evaluating its impact on either overall fertility or ovulatory infertility do not support a similar adverse effect.

Dairy and fertility. Consumption of three or more glasses of milk per day was protective of female fertility in an agricultural population in Wisconsin (124). In the NHS-II study no relation was found between total dairy intake and risk of ovulatory infertility, however anovulatory infertility was positively associated with low-fat and inversely associated with high-fat dairy intake (46). Similarly, the Biocycle study (125) reported that sporadic anovulation was more common among women consuming higher amounts of cream and yogurt. In the same study, total and low- and high-fat dairy food intakes were associated with a ~5% reduction in serum E_2 concentrations, and total and high-fat dairy food intakes were positively associated with serum LH.

Associations between dairy intake and fecundability in two separate cohorts of Danish and North American pregnancy planners (Snart Foraeldre and PRESTO, respectively) were inconsistent across cohorts. Total dairy and milk consumption was associated with increased fecundability in Denmark (where most milk is low-fat), whereas in the PRESTO cohort, there was little association between fecundability and dairy intake. In the latter, it was only among women younger than 30 years of age that cheese and high-dairy intake was associated with increased fecundability (126).

Dairy and ART outcomes. Finally, in a prospective cohort of women undergoing ART, dairy intake was positively associated with live birth only among women older than 35 years of age (127). Nevertheless, dairy food intake was not related to any of the evaluated intermediate outcomes (ie: response to gonadotropins, embryonic development, implantation and clinical pregnancy). The observed association did not differ between full-fat and low-fat dairy foods and did not appear to be driven by one single dairy food item (127).

Animal protein and reproductive outcomes. Regarding protein intake from nondairy sources and its effect on reproductive outcomes, available data point toward complex interactions between dietary protein and environmental contaminants. Ovarian antral follicle counts were not affected by either vegetable or animal protein intake from nondairy sources among women attending a fertility clinic (123). However, ovulation was negatively affected by increased meat intake among NHS-II participants. Intakes of fish, eggs, and processed meats did not seem to have an effect on anovulation, although increased vegetable protein intake seemed to decrease the risk (128). Similarly, blastocyst formation following ART was decreased among patients consuming more red meat. However, blastocyst formation was positively affected by fish consumption (129).

Regarding seafood consumption, most of the concerns relate to the fact that fish intake is the major pathway through

which humans are exposed to methyl mercury (130). Although early data indicated that mercury in fish may interfere with the endocrine system and impair fertility (131–133), recent prospective studies found no associations between mercury concentrations and in vitro fertilization end points (134) and time to pregnancy (135). Furthermore, strong relations between fish intake and shorter time to pregnancy (136), as well as between ω -3 PUFA and higher fecundability among non-fish oil supplement takers (99), support the benefits of consumption of fish with low mercury levels and high concentrations of ω -3 PUFA (99, 137). These findings were consistent with the most recent advice from the American College of Obstetricians and Gynecologists that pregnant women and those who may become pregnant are encouraged to eat two to three servings of a variety of fish per week, with no more than one serving per week of fish such as albacore tuna, and to avoid fish (e.g., bigeye tuna, king mackerel, swordfish) with the highest mercury concentrations (138).

Soy and reproductive outcomes. Given the available evidence suggesting that certain phytoestrogens may affect endocrine processes by influencing estrogen-dependent pathways (139–142), it is hypothesized that soy consumption by women attempting conception might affect fertility. Regarding a possible effect on ovarian aging, a limited number of studies evaluated a possible impact on serum FSH levels as a marker of ovarian reserve. Because the studies were small and heterogeneous and did not evaluate more precise markers of ovarian reserve, such as antimüllerian hormone and antral follicle counts, it is difficult to draw any definite conclusions (143–147). One would assume that if a deleterious effect of soy and phytoestrogens on ovarian aging exists, its consumption would accelerate the onset of menopause. From the available data, no such association between soy isoflavones and age at menopause has been observed (148–151). Regarding fecundity of populations with no history of infertility, women with the highest isoflavone intake participating in the retrospective Adventist Health Study were more likely to be childless, whereas no relation between soy intake and fecundity was noted in a prospective cohort of couples attempting pregnancy (152, 153). In women seeking fertility treatments, however, soy isoflavone supplements were associated with improvement in reproductive outcomes: increased live births after clomiphene administration (154) or higher endometrial thickness and ongoing pregnancy rates after intrauterine insemination (155) and in vitro fertilization (156). Similarly, dietary soy intake was positively related to the probability of live birth after ART among EARTH study participants (157).

Summary. Meat, fish, and dairy products are often vehicles for delivering environmental contaminants. To date, there is little evidence on the relationship of red meat and white meat to female fertility, and most literature did not disentangle the associations between these foods and environmental contaminants with reproductive outcomes. On the other hand, studies generally suggest that soy appears to have no adverse effects on female fertility, and may be beneficial for ART outcomes. Furthermore, despite concerns

regarding environmental contaminants, studies of fish intake and fertility suggest that the benefits of fish consumption may outweigh any risks posed by the environmental contaminants they may carry.

DIETARY PATTERNS

Dietary pattern analysis is an alternate and complementary approach to examine associations between overall diet quality and health outcomes (158).

Dietary Patterns and Reproductive Outcomes

In NHS-II, Chavarro et al. found that higher adherence to a “fertility diet” (consisting of high consumption of monounsaturated fat, vegetable protein, high-fat dairy, low-glycemic carbohydrates, multivitamins, and iron from plants and supplements) was associated with a lower risk of ovulatory infertility (159). Similarly, in a nested case-control study of university graduates in Spain, women with the highest adherence to the Mediterranean diet (characterized by high consumption of fruits and vegetables, fish and poultry, low-fat products, and olive oil) had lower odds of experiencing difficulties getting pregnant (160). Nonetheless, in the NHS-II cohort, adherence to a “fertility” or alternate Mediterranean diet, or to the alternate Healthy Eating Index 2010, was unrelated to risks of SAB (161). On the other hand, a retrospective study reported that higher frequency of fast food and lower frequency of fruit and vegetables intake was related to longer time to pregnancy (162).

Dietary Patterns and ART Outcomes

A favorable role of healthy dietary patterns on ART outcomes has been reported (163–165). In a cohort of 199 Dutch couples, increasing adherence to Dutch dietary recommendations (characterized by high intake of whole grains, fruits, vegetables, monounsaturated or polyunsaturated oils, meat or meat replacers, and fish) before conception was associated with higher probability of ongoing pregnancy (163). In a separate Dutch cohort of 161 couples, Vujkovic et al. found that maternal Mediterranean diet (high intake of vegetables and vegetable oils, fish, and legumes, low snack intake) but not “health conscious/low processed” diet (high intake of fruits, vegetables, legumes, whole grains, and fish, low intake of mayonnaise, snacks, and meat products) was associated with higher probability of pregnancy (164). Given that high intake of vegetable oil was dominant in the Mediterranean diet but not in the “health conscious/low processed” diet, these findings suggest that nutrients in vegetable oil, such as LA, may be responsible for this association. Similarly, in a recent study of young non-obese Greek women, a similar effect of adherence to Mediterranean dietary patterns and ART success was noted (165).

Summary

Although the definition of a healthy dietary pattern slightly varies across studies, these dietary patterns have remarkable overlaps in whole grains, fruits, vegetables, fish (rich in

long-chain ω -3 PUA), and olive oil (rich in MUFA), most of which have been shown to improve ART outcomes and chance of pregnancy. Nonetheless, dietary patterns were unrelated to risks of SAB, based on a large prospective study (161).

EMERGING CONCERNS

Foods are vehicles for delivering nutrients as well as nonnutritive constituent chemicals, which could be due to naturally occurring contaminants in the environment or artificially introduced during the processing, packaging, preparing, storage, and transportation of food. Well known examples include mercury in fish, pesticide residues in produce, and growth hormones, antibiotics, and polybrominated diphenyl ethers in meat. Emerging research has suggested that nutrients may interact with the toxicant in affecting specific health outcomes and vice versa. For example, in both human and nonhuman studies, intake of soy food and folates was found to be protective against the adverse reproductive effects of bisphenol-A (166–169). A recent study has shown that the presence of high pesticide residues in fruits and vegetables might modify the beneficial effects of fruits and vegetables on reproductive success (170). Because environmental contaminants are widely dispersed, future studies may need to consider coexposure to environmental contaminants (and mixtures of these contaminants) when investigating the associations between food/nutrients and health outcomes to better form dietary recommendations and guide action for general and susceptible populations.

CONCLUSION

Although much work remains, current evidence has accrued to support that diet could be a modifiable factor for female fecundity. According to existing data, intake of a folic acid supplement before and during pregnancy not only prevents birth defects, but also maintains pregnancy success. On the other hand, despite encouraging data in animal studies, little evidence supports beneficial effects of vitamin D on human fertility. There is considerable evidence suggesting that greater intake of ω -3 PUFA and lower intake of *trans* fats are associated with shorter time to pregnancy and better ART outcomes, whereas the effect from other fatty acids (such as ω -6 PUFA, SFA, and MUFA) on female fecundity is less clear. Furthermore, studies generally suggest that soy appears to have no harm on female fertility, and little data are available to disentangle the relationship of dairy, meat, and fish intake, environmental contaminants, and reproductive outcomes. Finally, greater adherence to healthy dietary patterns favoring whole grains, fish, fruits, vegetables, and olive oils—most of which are aligned with the 2015–2020 Dietary Guidelines for Americans (171)—may not only improve overall health but boost fecundity. Because most evidence came from observational studies, well designed RCTs are essential to validate these findings, which can be translated into practice and solid recommendations for pregnancy planners.

REFERENCES

1. Thoma ME, McLain AC, Louis JF, King RB, Trumble AC, Sundaram R, et al. Prevalence of infertility in the United States as estimated by the current duration approach and a traditional constructed approach. *Fertil Steril* 2013;99:1324–13231.e1.
2. Wilcox AJ, Weinberg CR, O'Connor JF, Baird DD, Schlatterer JP, Canfield RE, et al. Incidence of early loss of pregnancy. *N Engl J Med* 1988;319:189–94.
3. Willett WC. Diet and health: what should we eat? *Science* 1994;264:532–7.
4. Willett WC, Stampfer MJ. Current evidence on healthy eating. *Annu Rev Public Health* 2013;34:77–95.
5. Ley SH, Hamdy O, Mohan V, Hu FB. Prevention and management of type 2 diabetes: dietary components and nutritional strategies. *Lancet* 2014;383:1999–2007.
6. Schulze MB, Hu FB. Primary prevention of diabetes: what can be done and how much can be prevented? *Annu Rev Public Health* 2005;26:445–67.
7. Ebisch IM, Thomas CM, Peters WH, Braat DD, Steegers-Theunissen RP. The importance of folate, zinc and antioxidants in the pathogenesis and prevention of subfertility. *Hum Reprod Update* 2007;13:163–74.
8. Jaroudi S, SenGupta S. DNA repair in mammalian embryos. *Mutat Res* 2007;635:53–77.
9. Kiefer JC. Epigenetics in development. *Dev Dyn* 2007;236:1144–56.
10. Houk VN, Oakley GP, Erickson JD, Mulinare J, James LM. Recommendations for the use of folic acid to reduce the number of cases of spina bifida and other neural tube defects 1992.
11. Czeizel AE, Dudas I, Metneki J. Pregnancy outcomes in a randomised controlled trial of periconceptional multivitamin supplementation. Final report. *Arch Gynecol Obstet* 1994;255:131–9.
12. Windham GC, Shaw GM, Todoroff K, Swan SH. Miscarriage and use of multi-vitamins or folic acid. *Am J Med Genet* 2000;90:261–2.
13. Hook EB, Czeizel AE. Can terathanasia explain the protective effect of folic acid supplementation on birth defects? *Lancet* 1997;350:513–5.
14. Wald N, Hackshaw A. Folic acid and prevention of neural-tube defects. *Lancet* 1997;350:665.
15. Wald NJ, Hackshaw AK. Folic acid and miscarriage: an unjustified link. *Am J Med Genet* 2001;98:204.
16. Balogun OO, da Silva Lopes K, Ota E, Takemoto Y, Rumbold A, Takegata M, et al. Vitamin supplementation for preventing miscarriage. *Cochrane Database Syst Rev* 2016:CD004073.
17. MRC Vitamin Study Research Group. Prevention of neural tube defects: results of the Medical Research Council Vitamin Study. *Lancet* 1991;338:131–7.
18. Central Technical Co-ordinating Unit and ICMRCentral Technical Co-ordinating Unit. Multicentric study of efficacy of periconceptional folic acid containing vitamin supplementation in prevention of open neural tube defects from India. *Indian J Med Res* 2000;112:206–11.
19. Gindler J, Li Z, Berry RJ, Zheng J, Correa A, Sun X, et al. Folic acid supplements during pregnancy and risk of miscarriage. *Lancet* 2001;358:796–800.
20. Vila-Nova C, Wehby GL, Queiros FC, Chakraborty H, Felix TM, Goco N, et al. Periconceptional use of folic acid and risk of miscarriage—findings of the Oral Cleft Prevention Program in Brazil. *J Perinat Med* 2013;41:461–6.
21. Gaskins AJ, Rich-Edwards JW, Hauser R, Williams PL, Gillman MW, Ginsburg ES, et al. Maternal prepregnancy folate intake and risk of spontaneous abortion and stillbirth. *Obstet Gynecol* 2014;124:23–31.
22. Hasan R, Olshan AF, Herring AH, Savitz DA, Siega-Riz AM, Hartmann KE. Self-reported vitamin supplementation in early pregnancy and risk of miscarriage. *Am J Epidemiol* 2009;169:1312–8.
23. Byrne J. Periconceptional folic acid prevents miscarriage in Irish families with neural tube defects. *Ir J Med Sci* 2011;180:59–62.
24. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Use of multivitamins, intake of B vitamins, and risk of ovulatory infertility. *Fertil Steril* 2008;89:668–76.
25. Gaskins AJ, Mumford SL, Chavarro JE, Zhang C, Pollack AZ, Wactawski-Wende J, et al. The impact of dietary folate intake on reproductive function

in premenopausal women: a prospective cohort study. *PLoS One* 2012;7:e46276.

26. Cueto HT, Riis AH, Hatch EE, Wise LA, Rothman KJ, Sorensen HT, et al. Folic acid supplementation and fecundability: a Danish prospective cohort study. *Eur J Clin Nutr* 2016;70:66–71.
27. Westphal LM, Polan ML, Trant AS. Double-blind, placebo-controlled study of Fertilityblend: a nutritional supplement for improving fertility in women. *Clin Exp Obstet Gynecol* 2006;33:205–8.
28. Thaler CJ, Budiman H, Ruebsamen H, Nagel D, Lohse P. Effects of the common 677C>T mutation of the 5,10-methylenetetrahydrofolate reductase (MTHFR) gene on ovarian responsiveness to recombinant follicle-stimulating hormone. *Am J Reprod Immunol* 2006;55:251–8.
29. Hecht S, Pavlik R, Lohse P, Noss U, Friese K, Thaler CJ. Common 677C>T mutation of the 5,10-methylenetetrahydrofolate reductase gene affects follicular estradiol synthesis. *Fertil Steril* 2009;91:56–61.
30. Gaskins AJ, Afeiche MC, Wright DL, Toth TL, Williams PL, Gillman MW, et al. Dietary folate and reproductive success among women undergoing assisted reproduction. *Obstet Gynecol* 2014;124:801–9.
31. Gaskins AJ, Chiu YH, Williams PL, Ford JB, Toth TL, Hauser R, et al. Association between serum folate and vitamin B-12 and outcomes of assisted reproductive technologies. *Am J Clin Nutr* 2015;102:943–50.
32. Haggarty P, McCallum H, McBain H, Andrews K, Duthie S, McNeill G, et al. Effect of B vitamins and genetics on success of in-vitro fertilisation: prospective cohort study. *Lancet* 2006;367:1513–9.
33. Murto T, Kallak TK, Hoas A, Altmae S, Salumets A, Nilsson TK, et al. Folic acid supplementation and methylenetetrahydrofolate reductase (MTHFR) gene variations in relation to in vitro fertilization pregnancy outcome. *Acta Obstet Gynecol Scand* 2014;94:65–71.
34. Murto T, Skoog Svanberg A, Yngve A, Nilsson TK, Altmae S, Wanggren K, et al. Folic acid supplementation and IVF pregnancy outcome in women with unexplained infertility. *Reprod Biomed Online* 2014;28:766–72.
35. Szymanski W, Kazdepka-Zieminska A. [Effect of homocysteine concentration in follicular fluid on a degree of oocyte maturity]. *Ginekol Pol* 2003;74:1392–6, Polish.
36. Ebisch IM, Peters WH, Thomas CM, Wetzels AM, Peer PG, Steegers-Theunissen RP. Homocysteine, glutathione and related thiols affect fertility parameters in the (sub)fertile couple. *Hum Reprod* 2006;21:1725–33.
37. Boxmeer JC, Macklon NS, Lindemans J, Beckers NG, Eijkemans MJ, Laven JS, et al. IVF outcomes are associated with biomarkers of the homocysteine pathway in monofollicular fluid. *Hum Reprod* 2009;24:1059–66.
38. Lerchbaum E, Obermayer-Pietsch B. Vitamin D and fertility: a systematic review. *Eur J Endocrinol* 2012;166:765–78.
39. Kwiecinski GG, Petrie GI, DeLuca HF. 1,25-Dihydroxyvitamin D₃ restores fertility of vitamin D-deficient female rats. *Am J Physiol* 1989;256:E483–7.
40. Johnson LE, DeLuca HF. Vitamin D receptor null mutant mice fed high levels of calcium are fertile. *J Nutr* 2001;131:1787–91.
41. Kovacs CS, Woodland ML, Fudge NJ, Friel JK. The vitamin D receptor is not required for fetal mineral homeostasis or for the regulation of placental calcium transfer in mice. *Am J Physiol Endocrinol Metab* 2005;289:E133–44.
42. Panda DK, Miao D, Tremblay ML, Sirois J, Farookhi R, Hendy GN, et al. Targeted ablation of the 25-hydroxyvitamin D 1alpha-hydroxylase enzyme: evidence for skeletal, reproductive, and immune dysfunction. *Proc Natl Acad Sci U S A* 2001;98:7498–503.
43. Yoshizawa T, Handa Y, Uematsu Y, Takeda S, Sekine K, Yoshihara Y, et al. Mice lacking the vitamin D receptor exhibit impaired bone formation, uterine hypoplasia and growth retardation after weaning. *Nat Genet* 1997;16:391–6.
44. Irani M, Merhi Z. Role of vitamin D in ovarian physiology and its implication in reproduction: a systematic review. *Fertil Steril* 2014;102:460–468 e3.
45. Anagnostis P, Karras S, Goulis DG. Vitamin D in human reproduction: a narrative review. *Int J Clin Pract* 2013;67:225–35.
46. Chavarro JE, Rich-Edwards JW, Rosner B, Willett WC. A prospective study of dairy foods intake and anovulatory infertility. *Hum Reprod* 2007;22:1340–7.
47. Moller UK, Streym S, Heckendorff L, Mosekilde L, Rejnmark L. Effects of 25OHD concentrations on chances of pregnancy and pregnancy outcomes: a cohort study in healthy Danish women. *Eur J Clin Nutr* 2012;66:862–8.
48. Somigliana E, Paffoni A, Lattuada D, Colciaghi B, Filippi F, la Vecchia I, et al. Serum levels of 25-hydroxyvitamin D and time to natural pregnancy. *Gynecol Obstet Invest* 2016;81:468–71.
49. Zhang H, Huang Z, Xiao L, Jiang X, Chen D, Wei Y. Meta-analysis of the effect of the maternal vitamin D level on the risk of spontaneous pregnancy loss. *Int J Gynaecol Obstet* 2017;138:242–9.
50. Flood-Nichols SK, Tinnemore D, Huang RR, Napolitano PG, Ippolito DL. Vitamin D deficiency in early pregnancy. *PLoS One* 2015;10:e0123763.
51. Schneuer FJ, Roberts CL, Guilbert C, Simpson JM, Algert CS, Khambalia AZ, et al. Effects of maternal serum 25-hydroxyvitamin D concentrations in the first trimester on subsequent pregnancy outcomes in an Australian population. *Am J Clin Nutr* 2014;99:287–95.
52. Zhou J, Su L, Liu M, Liu Y, Cao X, Wang Z, et al. Associations between 25-hydroxyvitamin D levels and pregnancy outcomes: a prospective observational study in southern China. *Eur J Clin Nutr* 2014;68:925–30.
53. Andersen LB, Jorgensen JS, Jensen TK, Dalgard C, Barington T, Nielsen J, et al. Vitamin D insufficiency is associated with increased risk of first-trimester miscarriage in the Odense Child Cohort. *Am J Clin Nutr* 2015;102:633–8.
54. Aydogmus S, Kelekci S, Aydogmus H, Eris S, Desdicioglu R, Yilmaz B, et al. High prevalence of vitamin D deficiency among pregnant women in a Turkish population and impact on perinatal outcomes. *J Matern Fetal Neonatal Med* 2015;28:1828–32.
55. Anifandis GM, Dafopoulos K, Messini CI, Chalvatzas N, Liakos N, Pournaras S, et al. Prognostic value of follicular fluid 25-OH vitamin D and glucose levels in the IVF outcome. *Reprod Biol Endocrinol* 2010;8:91.
56. Firouzabadi RD, Rahmani E, Rahsepar M, Firouzabadi MM. Value of follicular fluid vitamin D in predicting the pregnancy rate in an IVF program. *Arch Gynecol Obstet* 2014;289:201–6.
57. Garbedian K, Boggild M, Moody J, Liu KE. Effect of vitamin D status on clinical pregnancy rates following in vitro fertilization. *CMAJ Open* 2013;1:E77–82.
58. Ozkan S, Jindal S, Greenseed K, Shu J, Zeitlian G, Hickmon C, et al. Replete vitamin D stores predict reproductive success following in vitro fertilization. *Fertil Steril* 2010;94:1314–9.
59. Paffoni A, Ferrari S, Viganò P, Pagliardini L, Papaleo E, Candiani M, et al. Vitamin D deficiency and infertility: insights from in vitro fertilization cycles. *J Clin Endocrinol Metab* 2014;99:E2372–6.
60. Fabris A, Pacheco A, Cruz M, Puente JM, Fatemi H, Garcia-Velasco JA. Impact of circulating levels of total and bioavailable serum vitamin D on pregnancy rate in egg donation recipients. *Fertil Steril* 2014;102:1608–12.
61. Franasiak JM, Molinaro TA, Dubell EK, Scott KL, Ruiz AR, Forman EJ, et al. Vitamin D levels do not affect IVF outcomes following the transfer of euploid blastocysts. *Am J Obstet Gynecol* 2015;212:315, e1–6.
62. Fru K, Segal T, Cox J, Mumford S, Sharara F, Segars J. Replete vitamin D levels are associated with higher pregnancy rates and increased number of live births in autologous IVF cycles. *Fertil Steril* 2014;102:e277.
63. Polyzos NP, Anckaert E, Guzman L, Schiettecatte J, Van Landuyt L, Camus M, et al. Vitamin D deficiency and pregnancy rates in women undergoing single embryo, blastocyst stage, transfer (SET) for IVF/ICSI. *Hum Reprod* 2014;29:2032–40.
64. Rudick B, Ingles SA, Chung K, Stanczyk F, Paulson R, Bendikson K. Characterizing the influence of vitamin D levels on IVF outcomes. *Hum Reprod* 2012;27:3321–7.
65. Rudick BJ, Ingles SA, Chung K, Stanczyk FZ, Paulson RJ, Bendikson KA. Influence of vitamin D levels on in vitro fertilization outcomes in donor-recipient cycles. *Fertil Steril* 2014;101:447–52.
66. Chu J, Gallos I, Tobias A, Tan B, Eapen A, Coomarasamy A. Vitamin D and assisted reproductive treatment outcome: a systematic review and meta-analysis. *Hum Reprod* 2018;33:65–80.
67. Pal L, Zhang H, Williams J, Santoro NF, Diamond MP, Schlaff WD, et al. Vitamin D status relates to reproductive outcome in women with polycystic ovary syndrome: secondary analysis of a multicenter randomized controlled trial. *J Clin Endocrinol Metab* 2016;101:3027–35.

68. Abadia L, Gaskins AJ, Chiu YH, Williams PL, Keller M, Wright DL, et al. Serum 25-hydroxyvitamin D concentrations and treatment outcomes of women undergoing assisted reproduction. *Am J Clin Nutr* 2016;104:729–35.
69. Aleyasin A, Hosseini MA, Mahdavi A, Safdarian L, Fallahi P, Mohajeri MR, et al. Predictive value of the level of vitamin D in follicular fluid on the outcome of assisted reproductive technology. *Eur J Obstet Gynecol Reprod Biol* 2011;159:132–7.
70. Neville G, Martyn F, Kilbane M, O'Riordan M, Wingfield M, McKenna M, et al. Vitamin D status and fertility outcomes during winter among couples undergoing in vitro fertilization/intracytoplasmic sperm injection. *Int J Gynaecol Obstet* 2016;135:172–6.
71. Aflatoonian A, Arabjehani F, Eftekhar M, Sayadi M. Effect of vitamin D insufficiency treatment on fertility outcomes in frozen-thawed embryo transfer cycles: a randomized clinical trial. *Iran J Reprod Med* 2014;12:595–600.
72. Asadi M, Matin N, Frootan M, Mohamadpour J, Qorbani M, Tanha FD. Vitamin D improves endometrial thickness in PCOS women who need intrauterine insemination: a randomized double-blind placebo-controlled trial. *Arch Gynecol Obstet* 2014;289:865–70.
73. Blaak EE. Carbohydrate quantity and quality and cardio-metabolic risk. *Curr Opin Clin Nutr Metab Care* 2016;19:289–93.
74. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. A prospective study of dietary carbohydrate quantity and quality in relation to risk of ovulatory infertility. *Eur J Clin Nutr* 2009;63:78–86.
75. Douglas CC, Norris LE, Oster RA, Darnell BE, Azziz R, Gower BA. Difference in dietary intake between women with polycystic ovary syndrome and healthy controls. *Fertil Steril* 2006;86:411–7.
76. Eslamian G, Baghestani AR, Eghtesad S, Hekmatdoost A. Dietary carbohydrate composition is associated with polycystic ovary syndrome: a case-control study. *J Hum Nutr Diet* 2017;30:90–7.
77. Altieri P, Cavazza C, Pasqui F, Morselli AM, Gambineri A, Pasquali R. Dietary habits and their relationship with hormones and metabolism in overweight and obese women with polycystic ovary syndrome. *Clin Endocrinol (Oxf)* 2013;78:52–9.
78. Douglas CC, Gower BA, Darnell BE, Ovalle F, Oster RA, Azziz R. Role of diet in the treatment of polycystic ovary syndrome. *Fertil Steril* 2006;85:679–88.
79. Gower BA, Chandler-Laney PC, Ovalle F, Goree LL, Azziz R, Desmond RA, et al. Favourable metabolic effects of a eucaloric lower-carbohydrate diet in women with PCOS. *Clin Endocrinol (Oxf)* 2013;79:550–7.
80. Mehrabani HH, Salehpour S, Amiri Z, Farahani SJ, Meyer BJ, Tahbaz F. Beneficial effects of a high-protein, low-glycemic-load hypocaloric diet in overweight and obese women with polycystic ovary syndrome: a randomized controlled intervention study. *J Am Coll Nutr* 2012;31:117–25.
81. Sjaarda LA, Schisterman EF, Schliep KC, Plowden T, Zarek SM, Yeung E, et al. Dietary Carbohydrate intake does not impact insulin resistance or androgens in healthy, eumenorrhic women. *J Clin Endocrinol Metab* 2015;100:2979–86.
82. Agarwal A, Aponte-Mellado A, Premkumar BJ, Shaman A, Gupta S. The effects of oxidative stress on female reproduction: a review. *Reprod Biol Endocrinol* 2012;10:49.
83. Adlercreutz H. Lignans and human health. *Crit Rev Clin Lab Sci* 2007;44:483–525.
84. Gaskins AJ, Chiu YH, Williams PL, Keller MG, Toth TL, Hauser R, et al. Maternal whole grain intake and outcomes of in vitro fertilization. *Fertil Steril* 2016;105:1503–15010 e4.
85. Gann PH, Chatterton RT, Gapstur SM, Liu K, Garside D, Giovanazzi S, et al. The effects of a low-fat/high-fiber diet on sex hormone levels and menstrual cycling in premenopausal women: a 12-month randomized trial (the Diet and Hormone Study). *Cancer* 2003;98:1870–9.
86. Woods MN, Barnett JB, Spiegelman D, Trail N, Hertzmark E, Longcope C, et al. Hormone levels during dietary changes in premenopausal African-American women. *J Natl Cancer Inst* 1996;88:1369–74.
87. Rose DP, Goldman M, Connolly JM, Strong LE. High-fiber diet reduces serum estrogen concentrations in premenopausal women. *Am J Clin Nutr* 1991;54:520–5.
88. Bagga D, Ashley JM, Geoffrey SP, Wang HJ, Barnard RJ, Korenman S, et al. Effects of a very low fat, high fiber diet on serum hormones and menstrual function. Implications for breast cancer prevention. *Cancer* 1995;76:2491–6.
89. Goldin BR, Woods MN, Spiegelman DL, Longcope C, Morrill-LaBrode A, Dwyer JT, et al. The effect of dietary fat and fiber on serum estrogen concentrations in premenopausal women under controlled dietary conditions. *Cancer* 1994;74:1125–31.
90. Goldin BR, Adlercreutz H, Gorbach SL, Warram JH, Dwyer JT, Swenson L, et al. Estrogen excretion patterns and plasma levels in vegetarian and omnivorous women. *N Engl J Med* 1982;307:1542–7.
91. Gaskins AJ, Mumford SL, Zhang C, Wactawski-Wende J, Hovey KM, Whitcomb BW, et al. Effect of daily fiber intake on reproductive function: the Biocycle study. *Am J Clin Nutr* 2009;90:1061–9.
92. Sturmev RG, Reis A, Leese HJ, McEvoy TG. Role of fatty acids in energy provision during oocyte maturation and early embryo development. *Reprod Domest Anim* 2009;44(Suppl 3):50–8.
93. Norwitz ER, Schust DJ, Fisher SJ. Implantation and the survival of early pregnancy. *N Engl J Med* 2001;345:1400–8.
94. Lefevre M, Lovejoy JC, Smith SR, Delany JP, Champagne C, Most MM, et al. Comparison of the acute response to meals enriched with cis- or trans-fatty acids on glucose and lipids in overweight individuals with differing FABP2 genotypes. *Metabolism* 2005;54:1652–8.
95. Kaipia A, Chun SY, Eisenhauer K, Hsueh AJ. Tumor necrosis factor-alpha and its second messenger, ceramide, stimulate apoptosis in cultured ovarian follicles. *Endocrinology* 1996;137:4864–70.
96. Mumford SL, Chavarro JE, Zhang C, Perkins NJ, Sjaarda LA, Pollack AZ, et al. Dietary fat intake and reproductive hormone concentrations and ovulation in regularly menstruating women. *Am J Clin Nutr* 2016;103:868–77.
97. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Dietary fatty acid intakes and the risk of ovulatory infertility. *Am J Clin Nutr* 2007;85:231–7.
98. Missmer SA, Chavarro JE, Malspeis S, Bertone-Johnson ER, Hornstein MD, Spiegelman D, et al. A prospective study of dietary fat consumption and endometriosis risk. *Hum Reprod* 2010;25:1528–35.
99. Wise LA, Wesselink AK, Tucker KL, Saklani S, Mikkelsen EM, Cueto H, et al. Dietary fat intake and fecundability in 2 preconception cohort studies. *Am J Epidemiol* 2018;187:60–74.
100. Al-Safi ZA, Liu H, Carlson NE, Chosich J, Harris M, Bradford AP, et al. Omega-3 fatty acid supplementation lowers serum FSH in normal weight but not obese women. *J Clin Endocrinol Metab* 2016;101:324–33.
101. Nehra D, Le HD, Fallon EM, Carlson SJ, Woods D, White YA, et al. Prolonging the female reproductive lifespan and improving egg quality with dietary omega-3 fatty acids. *Aging Cell* 2012;11:1046–54.
102. Chiu YH, Karmon AE, Gaskins AJ, Arvizu M, Williams PL, Souter I, et al. Serum omega-3 fatty acids and treatment outcomes among women undergoing assisted reproduction. *Hum Reprod* 2018;33:156–65.
103. Mirabi P, Chaichi MJ, Esmailzadeh S, Ali Jorsaraei SG, Bijani A, Ehsani M, et al. The role of fatty acids on ICSI outcomes: a prospective cohort study. *Lipids Health Dis* 2017;16:18.
104. Hammiche F, Vujkovic M, Wijburg W, de Vries JH, Macklon NS, Laven JS, et al. Increased preconception omega-3 polyunsaturated fatty acid intake improves embryo morphology. *Fertil Steril* 2011;95:1820–3.
105. Jungheim ES, Macones GA, Odem RR, Patterson BW, Moley KH. Elevated serum alpha-linolenic acid levels are associated with decreased chance of pregnancy after in vitro fertilization. *Fertil Steril* 2011;96:880–3.
106. Jungheim ES, Frolova AI, Jiang H, Riley JK. Relationship between serum polyunsaturated fatty acids and pregnancy in women undergoing in vitro fertilization. *J Clin Endocrinol Metab* 2013;98:E1364–8.
107. Moran LJ, Tsagareli V, Noakes M, Norman R. Altered preconception fatty acid intake is associated with improved pregnancy rates in overweight and obese women undertaking in vitro fertilisation. *Nutrients* 2016;8:E10.
108. Schaum J, Schuda L, Wu C, Sears R, Ferrario J, Andrews K. A national survey of persistent, bioaccumulative, and toxic (PBT) pollutants in the United States milk supply. *J Expo Anal Environ Epidemiol* 2003;13:177–86.

109. Liao C, Kannan K. Concentrations and profiles of bisphenol A and other bisphenol analogues in foodstuffs from the United States and their implications for human exposure. *J Agric Food Chem* 2013;61:4655–62.
110. Fraser AJ, Webster TF, McClean MD. Diet contributes significantly to the body burden of PBDEs in the general U.S. population. *Environ Health Perspect* 2009;117:1520–5.
111. Ganmaa D, Tezuka H, Enkhmaa D, Hoshi K, Sato A. Commercial cows' milk has uterotrophic activity on the uteri of young ovariectomized rats and immature rats. *Int J Cancer* 2006;118:2363–5.
112. Ganmaa D, Cui X, Feskanich D, Hankinson SE, Willett WC. Milk, dairy intake and risk of endometrial cancer: a 26-year follow-up. *Int J Cancer* 2012;130:2664–71.
113. Melnik BC, John SM, Carrera-Bastos P, Cordain L. The impact of cow's milk-mediated mTORC1-signaling in the initiation and progression of prostate cancer. *Nutr Metab (Lond)* 2012;9:74.
114. Jeong SH, Kang D, Lim MW, Kang CS, Sung HJ. Risk assessment of growth hormones and antimicrobial residues in meat. *Toxicol Res* 2010;26:301–13.
115. Hoppe C, Udam TR, Lauritzen L, Molgaard C, Juul A, Michaelsen KF. Animal protein intake, serum insulin-like growth factor I, and growth in healthy 2.5-y-old Danish children. *Am J Clin Nutr* 2004;80:447–52.
116. Norat T, Dossus L, Rinaldi S, Overvad K, Gronbaek H, Tjonneland A, et al. Diet, serum insulin-like growth factor-I and IGF-binding protein-3 in European women. *Eur J Clin Nutr* 2007;61:91–8.
117. Hoppe C, Molgaard C, Juul A, Michaelsen KF. High intakes of skimmed milk, but not meat, increase serum IGF-I and IGFBP-3 in eight-year-old boys. *Eur J Clin Nutr* 2004;58:1211–6.
118. Brinkman MT, Baglietto L, Krishnan K, English DR, Severi G, Morris HA, et al. Consumption of animal products, their nutrient components and postmenopausal circulating steroid hormone concentrations. *Eur J Clin Nutr* 2010;64:176–83.
119. Maruyama K, Oshima T, Ohyama K. Exposure to exogenous estrogen through intake of commercial milk produced from pregnant cows. *Pediatr Int* 2010;52:33–8.
120. Beasley JM, Gunter MJ, LaCroix AZ, Prentice RL, Neuhauser ML, Tinker LF, et al. Associations of serum insulin-like growth factor-I and insulin-like growth factor-binding protein 3 levels with biomarker-calibrated protein, dairy product and milk intake in the Women's Health Initiative. *Br J Nutr* 2014;111:847–53.
121. Bandyopadhyay S, Chakrabarti J, Banerjee S, Pal AK, Goswami SK, Chakravarty BN, et al. Galactose toxicity in the rat as a model for premature ovarian failure: an experimental approach readdressed. *Hum Reprod* 2003;18:2031–8.
122. Cramer DW, Xu H, Sahi T. Adult hypolactasia, milk consumption, and age-specific fertility. *Am J Epidemiol* 1994;139:282–9.
123. Souter I, Chiu YH, Batisis M, Afeiche MC, Williams PL, Hauser R, et al. The association of protein intake (amount and type) with ovarian antral follicle counts among infertile women: results from the EARTH prospective study cohort. *BJOG* 2017;124:1547–55.
124. Greenlee AR, Arbuckle TE, Chyow PH. Risk factors for female infertility in an agricultural region. *Epidemiology* 2003;14:429–36.
125. Kim K, Wactawski-Wende J, Michels KA, Plowden TC, Chaljub EN, Sjaarda LA, et al. Dairy food intake is associated with reproductive hormones and sporadic anovulation among healthy premenopausal women. *J Nutr* 2017;147:218–26.
126. Wise LA, Wesselink AK, Mikkelsen EM, Cueto H, Hahn KA, Rothman KJ, et al. Dairy intake and fecundability in 2 preconception cohort studies. *Am J Clin Nutr* 2017;105:100–10.
127. Afeiche MC, Chiu YH, Gaskins AJ, Williams PL, Souter I, Wright DL, et al. Dairy intake in relation to in vitro fertilization outcomes among women from a fertility clinic. *Hum Reprod* 2016;31:563–71.
128. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Protein intake and ovulatory infertility. *Am J Obstet Gynecol* 2008;198:210.e1–7.
129. Braga DP, Halpern G, Setti AS, Figueira RC, Iaconelli A Jr, Borges E Jr. The impact of food intake and social habits on embryo quality and the likelihood of blastocyst formation. *Reprod Biomed Online* 2015;31:30–8.
130. McDowell MA, Dillon CF, Osterloh J, Bolger PM, Pellizzari E, Fernando R, et al. Hair mercury levels in U.S. children and women of childbearing age: reference range data from NHANES 1999–2000. *Environ Health Perspect* 2004;112:1165–71.
131. Cole DC, Wainman B, Sanin LH, Weber JP, Muggah H, Ibrahim S. Environmental contaminant levels and fecundability among non-smoking couples. *Reprod Toxicol* 2006;22:13–9.
132. Choy CM, Lam CW, Cheung LT, Briton-Jones CM, Cheung LP, Haines CJ. Infertility, blood mercury concentrations and dietary seafood consumption: a case-control study. *BJOG* 2002;109:1121–5.
133. Buck GM, Vena JE, Schisterman EF, Dmochowski J, Mendola P, Sever LE, et al. Parental consumption of contaminated sport fish from Lake Ontario and predicted fecundability. *Epidemiology* 2000;11:388–93.
134. Wright DL, Afeiche MC, Ehrlich S, Smith K, Williams PL, Chavarro JE, et al. Hair mercury concentrations and in vitro fertilization (IVF) outcomes among women from a fertility clinic. *Reprod Toxicol* 2015;51:125–32.
135. Buck Louis GM, Sundaram R, Schisterman EF, Sweeney AM, Lynch CD, Gore-Langton RE, et al. Heavy metals and couple fecundity, the LIFE study. *Chemosphere* 2012;87:1201–7.
136. Gaskins AJ, Sundaram R, Buck Louis GM, Chavarro JE. Seafood intake, sexual activity, and time to pregnancy. *J Clin Endocrinol Metab* 2018;103:2680–8.
137. Hsi HC, Hsu YW, Chang TC, Chien LC. Methylmercury concentration in fish and risk-benefit assessment of fish intake among pregnant versus infertile women in Taiwan. *PLoS One* 2016;11:e0155704.
138. American College of Obstetricians and Gynecologists. ACOG Practice advisory: update on seafood consumption during pregnancy. 2017. Available at: <https://www.acog.org/Clinical-Guidance-and-Publications/Practice-Advisories/ACOG-Practice-Advisory-Seafood-Consumption-During-Pregnancy>.
139. Hwang CS, Kwak HS, Lim HJ, Lee SH, Kang YS, Choe TB, et al. Isoflavone metabolites and their in vitro dual functions: they can act as an estrogenic agonist or antagonist depending on the estrogen concentration. *J Steroid Biochem Mol Biol* 2006;101:246–53.
140. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 1998;139:4252–63.
141. Unfer V, Casini ML, Costabile L, Mignosa M, Gerli S, di Renzo GC. Endometrial effects of long-term treatment with phytoestrogens: a randomized, double-blind, placebo-controlled study. *Fertil Steril* 2004;82:145–8.
142. Amir AA, Kelly JM, Kleemann DO, Durmic Z, Blache D, Martin GB. Phyto-oestrogens affect fertilisation and embryo development in vitro in sheep. *Reprod Fertil Dev* 2018 Feb 16. doi: 10.1071/RD16481. [Epub ahead of print.]
143. Tsuji M, Tamai Y, Wada K, Nakamura K, Hayashi M, Takeda N, et al. Associations of intakes of fat, dietary fiber, soy isoflavones, and alcohol with levels of sex hormones and prolactin in premenopausal Japanese women. *Cancer Causes Control* 2012;23:683–9.
144. Duncan AM, Merz BE, Xu X, Nagel TC, Phipps WR, Kurzer MS. Soy isoflavones exert modest hormonal effects in premenopausal women. *J Clin Endocrinol Metab* 1999;84:192–7.
145. Brown BD, Thomas W, Hutchins A, Martini MC, Slavin JL. Types of dietary fat and soy minimally affect hormones and biomarkers associated with breast cancer risk in premenopausal women. *Nutr Cancer* 2002;43:22–30.
146. Maskarinec G, Williams AE, Inouye JS, Stanczyk FZ, Franke AA. A randomized isoflavone intervention among premenopausal women. *Cancer Epidemiol Biomarkers Prev* 2002;11:195–201.
147. Zittermann A, Geppert J, Baier S, Zehn N, Gouni-Berthold I, Berthold HK, et al. Short-term effects of high soy supplementation on sex hormones, bone markers, and lipid parameters in young female adults. *Eur J Nutr* 2004;43:100–8.
148. Nagata C, Wada K, Nakamura K, Tamai Y, Tsuji M, Shimizu H. Associations of physical activity and diet with the onset of menopause in Japanese women. *Menopause* 2012;19:75–81.
149. Nagata C, Shimizu H, Takami R, Hayashi M, Takeda N, Yasuda K. Serum concentrations of estradiol and dehydroepiandrosterone sulfate and soy product intake in relation to psychological well-being in peri- and postmenopausal Japanese women. *Metabolism* 2000;49:1561–4.

150. Dorjgochoo T, Kallianpur A, Gao YT, Cai H, Yang G, Li H, et al. Dietary and lifestyle predictors of age at natural menopause and reproductive span in the Shanghai Women's Health Study. *Menopause* 2008;15:924–33.
151. Nagel G, Altenburg HP, Nieters A, Boffetta P, Linseisen J. Reproductive and dietary determinants of the age at menopause in EPIC-Heidelberg. *Maturitas* 2005;52:337–47.
152. Jacobsen BK, Jaceldo-Siegl K, Knutsen SF, Fan J, Oda K, Fraser GE. Soy isoflavone intake and the likelihood of ever becoming a mother: the Adventist Health Study-2. *Int J Womens Health* 2014;6:377–84.
153. Mumford SL, Sundaram R, Schisterman EF, Sweeney AM, Barr DB, Rybak ME, et al. Higher urinary lignan concentrations in women but not men are positively associated with shorter time to pregnancy. *J Nutr* 2014;144:352–8.
154. Shahin AY, Ismail AM, Zahran KM, Makhlof AM. Adding phytoestrogens to clomiphene induction in unexplained infertility patients—a randomized trial. *Reprod Biomed Online* 2008;16:580–8.
155. Unfer V, Casini ML, Costabile L, Mignosa M, Gerli S, di Renzo GC. High dose of phytoestrogens can reverse the antiestrogenic effects of clomiphene citrate on the endometrium in patients undergoing intrauterine insemination: a randomized trial. *J Soc Gynecol Investig* 2004;11:323–8.
156. Unfer V, Casini ML, Gerli S, Costabile L, Mignosa M, di Renzo GC. Phytoestrogens may improve the pregnancy rate in in vitro fertilization-embryo transfer cycles: a prospective, controlled, randomized trial. *Fertil Steril* 2004;82:1509–13.
157. Vanegas JC, Afeiche MC, Gaskins AJ, Minguez-Alarcon L, Williams PL, Wright DL, et al. Soy food intake and treatment outcomes of women undergoing assisted reproductive technology. *Fertil Steril* 2015;103:749–755 e2.
158. Hu FB. Dietary pattern analysis: a new direction in nutritional epidemiology. *Curr Opin Lipidol* 2002;13:3–9.
159. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Diet and lifestyle in the prevention of ovulatory disorder infertility. *Obstet Gynecol* 2007;110:1050–8.
160. Toledo E, Lopez-del Burgo C, Ruiz-Zambrana A, Donazar M, Navarro-Blasco I, Martinez-Gonzalez MA, et al. Dietary patterns and difficulty conceiving: a nested case-control study. *Fertil Steril* 2011;96:1149–53.
161. Gaskins AJ, Rich-Edwards JW, Hauser R, Williams PL, Gillman MW, Penzias A, et al. Prepregnancy dietary patterns and risk of pregnancy loss. *Am J Clin Nutr* 2014;100:1166–72.
162. Grieger JA, Grzeskowiak LE, Bianco-Miotto T, Jankovic-Karasoulos T, Moran LJ, Wilson RL, et al. Pre-pregnancy fast food and fruit intake is associated with time to pregnancy. *Hum Reprod* 2018;33:1063–70.
163. Twigt JM, Bolhuis ME, Steegers EA, Hammiche F, van Inzen WG, Laven JS, et al. The preconception diet is associated with the chance of ongoing pregnancy in women undergoing IVF/ICSI treatment. *Hum Reprod* 2012;27:2526–31.
164. Vujkovic M, de Vries JH, Lindemans J, Macklon NS, van der Spek PJ, Steegers EA, et al. The preconception Mediterranean dietary pattern in couples undergoing in vitro fertilization/intracytoplasmic sperm injection treatment increases the chance of pregnancy. *Fertil Steril* 2010;94:2096–101.
165. Karayiannis D, Kontogianni MD, Mendorou C, Mastrominas M, Yiannakouris N. Adherence to the Mediterranean diet and IVF success rate among non-obese women attempting fertility. *Hum Reprod* 2018.
166. Chavarro JE, Minguez-Alarcon L, Chiu YH, Gaskins AJ, Souter I, Williams PL, et al. Soy intake modifies the relation between urinary bisphenol A concentrations and pregnancy outcomes among women undergoing assisted reproduction. *J Clin Endocrinol Metab* 2016;101:1082–90.
167. Minguez-Alarcon L, Gaskins AJ, Chiu YH, Souter I, Williams PL, Calafat AM, et al. Dietary folate intake and modification of the association of urinary bisphenol A concentrations with in vitro fertilization outcomes among women from a fertility clinic. *Reprod Toxicol* 2016;65:104–12.
168. Muhlhauser A, Susiarjo M, Rubio C, Griswold J, Gorence G, Hassold T, et al. Bisphenol A effects on the growing mouse oocyte are influenced by diet. *Biol Reprod* 2009;80:1066–71.
169. Dolinoy DC, Huang D, Jirtle RL. Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development. *Proc Natl Acad Sci U S A* 2007;104:13056–61.
170. Chiu YH, Williams PL, Gillman MW, Gaskins AJ, Minguez-Alarcon L, Souter I, et al. Association between pesticide residue intake from consumption of fruits and vegetables and pregnancy outcomes among women undergoing infertility treatment with assisted reproductive technology. *JAMA Intern Med* 2018;178:17–26.
171. Millen BE, Abrams S, Adams-Campbell L, Anderson CA, Brenna JT, Campbell WW, et al. The 2015 Dietary Guidelines Advisory Committee scientific report: development and major conclusions. *Adv Nutr* 2016;7:438–44.